

STUDY PROTOCOL

The race-based stress reduction intervention (RiSE) study on African American women in NYC and Chicago: Design and methods for complex genomic analysis

Jacquelyn Y. Taylor¹, Alexandria Jones-Patten¹, Laura Prescott¹, Stephanie Potts-Thompson^{1*}, Cara Joyce², Bamidele Tayo², Karen Saban³

1 Center for Research on People of Color, Columbia University School of Nursing, New York, New York, United States of America, **2** Parkinson School of Health Sciences and Public Health, Loyola University Chicago, Maywood, Illinois, United States of America, **3** Marcella Niehoff School of Nursing, Center for Translational Research and Education, Loyola University Chicago, Maywood, Illinois, United States of America

* sp3879@cumc.columbia.edu



OPEN ACCESS

Citation: Taylor JY, Jones-Patten A, Prescott L, Potts-Thompson S, Joyce C, Tayo B, et al. (2024) The race-based stress reduction intervention (RiSE) study on African American women in NYC and Chicago: Design and methods for complex genomic analysis. *PLoS ONE* 19(4): e0295293. <https://doi.org/10.1371/journal.pone.0295293>

Editor: Aymen Ahmed, Dow University of Health Sciences, PAKISTAN

Received: May 24, 2023

Accepted: November 19, 2023

Published: April 10, 2024

Copyright: © 2024 Taylor et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: No datasets were generated or analysed during the current study. All relevant data from this study will be made available upon study completion.

Funding: This work has been partially funded by the following: National Institute on Aging R01AG081251 to Drs. Saban and Taylor; and National Heart, Lung, and Blood Institute T32HL007343 to Drs. Taylor and Jones-Patten. The funders did not and will not have a role in

Abstract

RiSE study aims to evaluate a race-based stress-reduction intervention as an effective strategy to improve coping and decrease stress-related symptoms, inflammatory burden, and modify DNA methylation of stress response-related genes in older AA women. This article will describe genomic analytic methods to be utilized in this longitudinal, randomized clinical trial of older adult AA women in Chicago and NYC that examines the effect of the RiSE intervention on DNAm pre- and post-intervention, and its overall influence on inflammatory burden. Salivary DNAm will be measured at baseline and 6 months following the intervention, using the Oragene-DNA kit. Measures of perceived stress, depressive symptoms, fatigue, sleep, inflammatory burden, and coping strategies will be assessed at 4 time points including at baseline, 4 weeks, 8 weeks, and 6 months. Genomic data analysis will include the use of pre-processed and quality-controlled methylation data expressed as beta (β) values. Association analyses will be performed to detect differentially methylated sites on the targeted candidate genes between the intervention and non-intervention groups using the $\Delta\beta$ (changes in methylation) with adjustment for age, health behaviors, early life adversity, hybridization batch, and top principal components of the probes as covariates. To account for multiple testing, we will use FDR adjustment with a corrected p-value of <0.05 regarded as statistically significant. To assess the relationship between inflammatory burden and $\Delta\beta$ among the study samples, we will repeat association analyses with the inclusion of individual inflammation protein measures. ANCOVA will be used because it is more statistically powerful to detect differences.

study design, data collection and analysis, the decision to publish, or the preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Introduction

Impact of psychosocial stress on DNAm

Changes in DNA methylation (DNAm) may be some of the earliest cellular events in disease onset [1], such aberrant changes have been linked to inflammatory burden [2, 3] and a broad range of diseases including cardiometabolic disease (CMD) [4, 5]. Compelling evidence demonstrates that DNAm is associated with early life adversity [6–8], social stress [9], and history of trauma [10, 11]. Our recent epigenome-wide association study [12] revealed that greater levels of perceived discrimination are associated with decreased DNAm at seven CpG sites linked to inflammatory disease-associated genes in a sample of African American (AA) women [12]. Importantly, DNAm associated with psychological stress is malleable [13], making it a prime target for psychobehavioral interventions [13–16]. To date, only a few studies have explored the impact of psychobehavioral interventions on changes in DNAm of stress response-related genes with the majority focusing on mindfulness interventions. In a small study of veterans with post-traumatic stress disorder who participated in a mindfulness-based stress reduction program, investigators found that, in addition to improvements in trauma symptoms, participants also had an increase in methylation of the FKBP5, a gene involved in modulating glucocorticoid receptor activity [17, 18]. Others have found a decrease in methylation of SLC6A4, a gene related to serotonin transport, following participation in a three-month mindfulness program [19]. In a small randomized, wait-listed control trial, women who participated in an 8-week yoga intervention had reduced methylation of the TNF region as compared to women in the control group [20]. Changes in DNAm have also been associated with tai chi [21], psychotherapy [22], and meditation [23]. The most commonly identified genes associated with DNAm changes following a psychobehavioral intervention include SLC6A4 [19], FKBP5 [22, 24], and BDNF [25, 26].

Importance of genomics

Genomics has brought significant contributions to cardiovascular disease (CVD) research [27], particularly among AAs, including the combined effects of genetics and cigarette smoking on systolic BP [27] and the effects of body mass index (BMI) on DNAm [28]. DNAm is one type of epigenetic process that modulates gene expression by adding or removing methyl groups to DNA in response to the environment. Studies demonstrate that hyper or hypo methylation of genes due to chronic stressors, including racism and discrimination, are significantly associated with CMD risk [29–36]. Emerging evidence demonstrates that psychobehavioral interventions may modify the methylation of stress response-related genes (e.g., TNF, SLC6A4, FKBP5) [15, 21], potentially buffering the impact of psychological stress at the molecular level. However, few studies have examined the impact of a psychobehavioral intervention on changes in DNAm and none have addressed chronic stress in older AA women.

Here we will describe the methods that will be utilized in a randomized clinical trial of AA Women in Chicago and New York City that examines the effect of the RiSE intervention on DNAm pre and post-intervention, and its overall influence on cardiometabolic risk outcomes over time. We will use a targeted candidate gene approach by selecting 20 stress-related candidate genes (and their corresponding CpG sites), assessing DNAm pre and post-RiSE and their association with the outcome measures of cardiometabolic risk (perceived stress, depressive symptoms, and fatigue and sleep disturbance). See **Table 1** and **Fig 1**.

Resilience, Stress, and Ethnicity (RiSE) [37–39], is a group-based, 8-week intervention that integrates cognitive-behavioral strategies [40] focused on the biopsychosocial impact of racism [41, 42], racial identity development [43–45], and empowerment [46]. The conceptual model

Table 1. Sample size estimation based on preliminary date. *Assumes n = 125 per group complete cases and $\alpha = 0.05$; r-squared of 5 covariates is 0.30, **The smallest mean difference necessary to ensure > 80% power, assuming pooled SD observed in the pilot.

Outcome evaluated at 6 months	RiSE Mean	Control Mean	\Delta Means	Pooled SD	Effect size	Power*	Mean difference for 80% power**
Observed: Pilot Data @ 8 weeks							
WCQ-Active coping	23.76	24.77	1.01	8.24	0.06	21%	2.45
WCQ-Avoidance coping	8.14	14.38	6.24	5.25	0.59	>99%	1.56
WCQ-minimize situation	11.21	14.04	2.83	4.36	0.32	>99%	1.30
DASS-21: Depression	1.45	3.13	1.68	3.09	0.27	>99%	0.92
DASS-21: Anxiety	2.39	3.31	0.92	3.41	0.13	72%	1.02
DASS-21: Stress	4.45	5.90	1.45	4.05	0.18	92%	1.20
TNF-a	0.95	1.00	0.05	0.36	0.07	26%	0.11
High-sensitivity C-reactive protein (hsCRP)	3.83	7.24	3.41	6.30	0.27	>99%	1.88

<https://doi.org/10.1371/journal.pone.0295293.t001>

for the study is based on allostatic load theory [47] which posits that accumulating stress or “wear and tear” results in negative physiological consequences. See Fig 2.

Based on our preliminary work, we anticipate that participation in RiSE will reduce psychological distress and inflammation. RiSE is a stress reduction intervention that has been shown

	STUDY PERIOD				
	Enrolment	Allocation	Post-allocation		Close-out
	$-t_1$	t_0	t_1	t_2	t_3
ENROLMENT					
Eligibility screen	X				
Informed consent		X			
1 st DNA Sample		X			
Allocation		X			
INTERVENTIONS					
Health Measures		X	X	X	X
ACASI		X	X	X	X
2 nd DNA Sample					X
ASSESSMENTS					
Post-Evaluation					X

Fig 1. Study period. Schedule of enrollment, interventions, and assessments.

<https://doi.org/10.1371/journal.pone.0295293.g001>

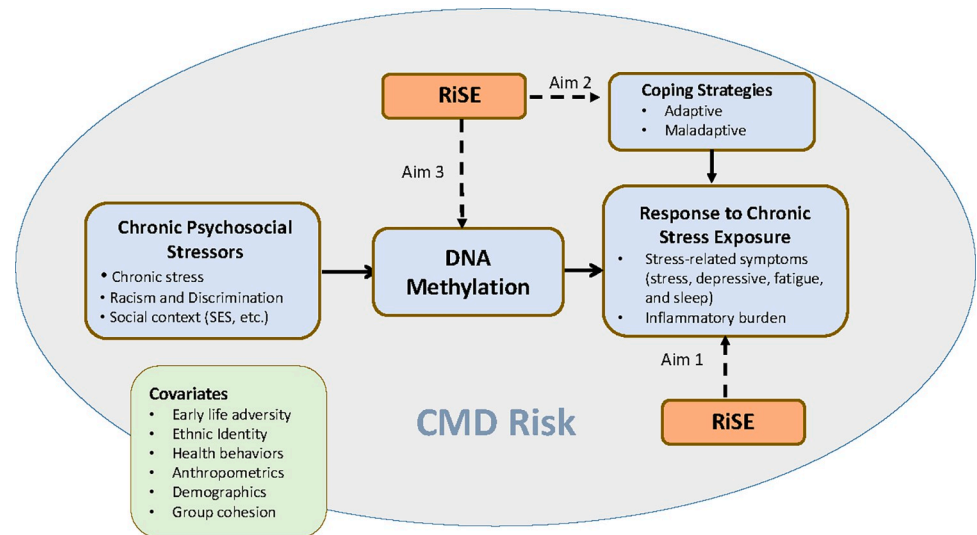


Fig 2. Conceptual model.

<https://doi.org/10.1371/journal.pone.0295293.g002>

to be effective in improving coping among AA women [48]. Further, in a targeted candidate gene analysis and epigenomic study in AA women, we found that DNAm patterns of four genes related to the regulation of blood pressure (BP) were associated with stress overload, problem-solving coping, social support coping, and avoidance coping [49], although these associations were not significant after correcting for multiple testing. Others have reported associations between DNAm and stress-related symptoms such as perceived stress, depressive symptoms, fatigue, and sleep [50–52].

Psychosocial stressors, such as discrimination, racism, and social context (e.g., socioeconomic status), when chronic and relentless, can impact the DNAm of stress-response related genes [12, 53], altering the expression of genes that regulate inflammatory response to stress [53]. Based on our preliminary data as well as evidence from other stress reduction interventions [54, 55], we propose that RiSE will improve stress-related symptoms and decrease inflammation (e.g., CRP, IL-6, TNF α , IL-1 β , and IFN- γ) [38].

We will use a targeted candidate gene approach by selecting 20 genes (and their corresponding CpG sites) that are associated with our outcome measures to assess DNAm modifications following RiSE (Table 2).

Our candidate genes were selected based on those consistently associated with stress response in the literature (see Table 2). Several studies have demonstrated that DNAm can be modified within weeks to months following changes in environmental factors [79–81]. In those previous studies of longitudinal change of DNAm, a large variation of inter-individual differences over time has been observed [82]. Therefore, our proposed 6-month follow-up not only provides a reasonable time frame to investigate the longitudinal change in DNAm but also links to socio-behavioral stressors among AA women, which have not been previously studied.

Inflammatory markers

Stress is related to low-grade inflammation, as measured by C-reactive protein (CRP) and proinflammatory cytokines such as interleukin-6 (IL-6), tumor necrosis factor (TNF- α), interleukin-1 beta (IL-1 β), and interferon-gamma (IFN- γ) [83–88] as well as increased CMD risk [89]. We will assess the relationship between inflammatory burden IL 6, TNF, IL-1B, IFNG,

Table 2. Stress-related candidate genes and functions.

Gene	Function	Gene	Function
FKBP5	Encodes FK506-binding protein 51. Pleiotropic effects on stress and inflammation, associated with CMD [56]	PAC1	Regulates PACAP which is linked to the stress response [57]
SLC6A4	Codes for the serotonin transporter and is associated with depression [58]	5-HTT	Regulation of serotonin transporter protein which is associated with depression and stress [59]
BDNF	Regulates neurogenesis in brain. Alterations of BDNF associated with stress and plays role in regulating body weight [60, 61]	CRF	Regulates homeostasis and neuroendocrine response to stress [62]
TNF	Encodes proinflammatory cytokine. Associated with atherosclerosis [4, 63]	TLR1	Mediates cytokine secretion and inflammatory response to stress [62]
OXTR	Encodes oxytocin receptor and is associated with range of psychological responses [64]	COMT	Instructs enzyme that regulates neurotransmitters associated with stress [65]
IL-6	Regulates proinflammatory cytokine IL-6 [66]	OPRM1	Provides instructions for making mu opioid receptor which is associated with PTSD [67]
NR3C1	Encodes glucocorticoid receptor associated with stress response and CMD [68–70]	KITLG	Inhibitory role on natural killer cells. Associated with cortisol stress reactivity [71]
AVP	Encodes member of vasopressin/oxytocin family. Plays role in glucocorticoid signaling [62]	DNMT-1	Provides instructions for making DNA methyltransferase 1 which is associated with stress response as well as CMD [4, 72, 73]
GABA	Provide instructions for making GABAA receptor protein which produces calming effect [74]	HSD11β2	Implicated in glucocorticoid regulation and response to stress. Associated with CVD [75, 76]
MAOA	Regulates monoamine oxidase A that is related to psychosocial stress [77]	ACE	Regulates angiotensin converting enzyme which is associated with depression and CVD [78]

<https://doi.org/10.1371/journal.pone.0295293.t002>

and CRP and change in methylation among the study samples. See the ‘Genomic Data Analysis’ section below for a description of the analysis with the inclusion of individual inflammatory protein measures.

Methods

This research was approved by Loyola University’s Institutional Review Board (IRB) Protocol number LU214133 WCG IRB #20230283. This study is registered through ClinicalTrials.gov, registration number: NCT05902741. It is estimated that the recruitment of participants will begin summer of 2023.

Procedures

Detailed descriptions of this study have been previously discussed [90]. Briefly, this convenience sample will include 300 self-identified AA women residing in either the Chicagoland (n = 150) or New York City (n = 150) area between the ages of 50 and 75 years with risk factors for CMD [91]. Participants will be randomized to either an 8-week RiSE intervention or a Health Education Program (HEP) (attention control). Potential participants will be pre-screened for eligibility over the phone and will be followed by an appointment for further screening if the potential participant is eligible. After collecting informed consent for obtaining inflammatory markers and DNAm samples, the samples will be stored in a lab for processing and frozen for later batch analysis. DNAm samples will be stored at 4°C. Specimens will be shipped overnight on dry ice to respective labs for analysis per best practice guidelines [92]. Individuals will contribute data for up to four-time points (baseline, 4 weeks (mid-intervention), 8 weeks (completion of intervention), and 6 months post-intervention). Data will be collected from participants using an Audio Computer-Assisted Self-Interview system (ACASI) [93]. Perceived stress, depression, fatigue, and sleep disturbance symptoms will be analyzed via the Perceived Stress Scale (PSS-10) [94], Patient Health Questionnaire 9 (PHQ 9) [95], NIH PROMIS Fatigue Short Form 8a [96], and NIH PROMIS Short Form v1.0 [97].

DNA sample collection

Salivary DNAm will be measured at baseline and 6 months following the intervention. Saliva was selected as the sample of choice as it correlates well with DNAm in blood [98–100], is more similar to patterns of DNAm in the brain as compared to blood [101–102], associates with CMD indices [103], and correlates with frontolimbic brain function [104, 105] and therefore, potentially offering distinct opportunities for studies considering psychological measures function [104, 105].

Participants will collect saliva samples using the Oragene-DNA (OG-510) kit that requires them to spit saliva into a tube up to the 1mL mark. Taylor et al. have described details of DNA collection and analysis procedures elsewhere [106]. Samples will be refrigerated at 4 C° in a lab until DNA extraction and analysis are completed. All analysis of DNA extraction will be done at Columbia University under the guidance of Dr. Taylor. Samples collected at Loyola University will be shipped at ambient temperature to Columbia University for processing and analysis. All tubes and plates that contain an individual's DNA will be labeled with a barcode to ensure precise sample tracking and recorded in the laboratory's computerized freezer inventory via barcodes upon arrival at the laboratory.

Genotyping

The Illumina Infinium Global Diversity Array (GDA) will be used to assess the targeted candidate genes (Table 2) and for ancestry informative markers (AIMS). AIMS are included on the GDA chips for multiethnic populations which is suited for our sample of AA women. The GDA combines highly optimized multiethnic, genome-wide content with curated clinical research variants. We selected the GDA chip because it has expanded coverage of SNPs for multiethnic populations [106].

DNAm analysis

The Illumina Infinium Methylation EPIC v2.0 (900K) BeadChip will be used for examining the methylation of targeted candidate genes [107]. We will apply standard quality control measures including accounting for cell type, batch, and plate effects. This BeadChip directly quantifies DNAm at 936,866 CpG dinucleotides, giving near complete coverage of known genes [107–109]. We will perform hybridization on a per-sample basis. The Infinium arrays are well annotated for CpG dinucleotides in CpG island and non-CpG island promoters, shore regions, coding regions, repetitive elements, miRNA promoter regions, FANTOM5 enhancers, ENCODE open chromatin and enhancers, and DNase hypersensitivity sites and include 91.1% of the loci from the Human-Methylation450 BeadChip. DNAm is determined at each of the CpG sites on the 935K array by measuring the fluorescent signals from the M (methylated) and U (unmethylated) probes specific for each site included in the array, covering approximately 99% of all RefSeq genes and 96% of CpG islands [110]. We will confirm DNAm by methylation-specific polymerase chain reaction (PCR) and by bisulfite sequencing [111]. We may explore the use of additional methylation data in the future to identify additional novel individual genes with abnormal gain and/or loss of CpG methylation associated with social and CMD features for future studies. Additional data from the BeadChip will be retained for future analyses as new information regarding epigenomic differences associated with CMD risk and social factors are described in the literature.

Data analytic plan

Power analysis. We will focus primarily on the tested CpG sites that are true positives, i.e., detected CpG sites with meaningful differential DNAm. There is currently no standard

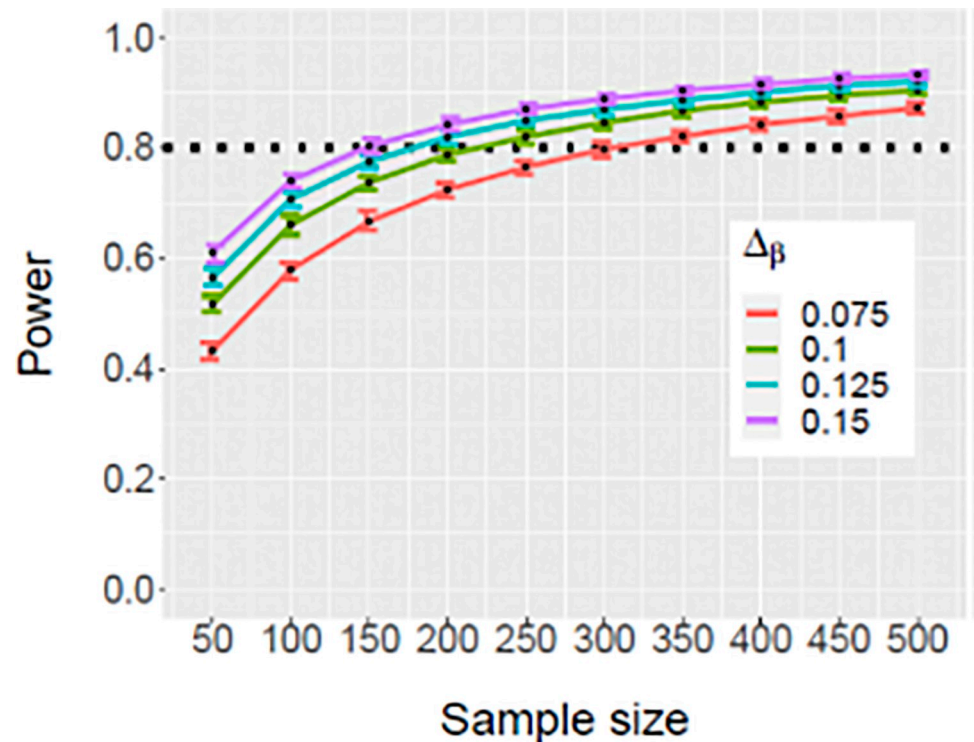


Fig 3. Plot of mean simulated power with 95 percentile (2.5 & 97.5%) for select DNAm differences.

<https://doi.org/10.1371/journal.pone.0295293.g003>

threshold for meaningful differential DNAm, though previous studies have used thresholds of 5–20% [112] for meaningful differential DNAm, and for this study, we use a conservative threshold of 10%. Using the R package ‘pwrEWAS’ [113], we performed simulations to evaluate sample sizes required to detect effect sizes (percent differences) of 7.5%, 10%, 12.5%, and 15% in CpG-specific methylation across 4000 CpGs on our 20 candidate genes with at least 80% power. We need 200 total subjects (150 per group) to detect a difference of at least 10% with at least 80% power across a set of CpG sites (Fig 3). Our simulations also indicated that the power to detect at least one differentially methylated CpG out of the set of 4000 is above 85%.

Data quality and integrity will be ensured before the conduct of statistical analysis. Prior power analyses for each planned analysis were conducted to ensure that each analysis was appropriately powered. All post-hoc tests will be controlled for multiple comparisons (i.e., Type I error). ANCOVA will also be used because it is better at controlling for chance imbalances in randomized groups at baseline, avoids regression to the mean issues, and is generally more statistically powerful in detecting differences [114].

Genomic data analysis. Using the pre-processed and quality-controlled methylation data expressed as beta values which are the percentage methylation at each CpG site at specific targeted candidate genes, we will determine changes in methylation ($\Delta\beta$) at every site as the difference between methylation at baseline and methylation at post-intervention. Negative $\Delta\beta$ indicates a decrease in methylation from baseline to post-intervention, and a positive value indicates an increase in methylation from baseline to post-intervention. Association analyses will be performed to detect differentially methylated sites on the targeted candidate genes between the intervention and non-intervention groups using the $\Delta\beta$ with adjustment for age, health behaviors, early life adversity, hybridization batch, and top principal components of the

probes as covariates. To adjust for multiple testing, we will use False Discovery Rate (FDR) adjustment with an FDR of <0.05 regarded as statistically significant. All analyses will be performed in R software using the Bioconductor R package 'limma' [115]. To assess the relationship between inflammatory burden (IL-6, TNF- α , IL-1 β , IFN- γ , CRP) and $\Delta\beta$ among the study samples, we will repeat the above association analyses with the inclusion of individual inflammatory protein measure.

Discussion

Potential pitfalls, alternative approaches

Given the longitudinal nature of this study and the significant commitment necessary to participate in the study, attrition is an inherent potential issue. In our preliminary work, we found an 11% attrition rate with a similar design. For this study, we will conservatively over-sample by 20% to accommodate potential attrition to achieve $n = 250$ participants with complete data at 6 months. We will minimize attrition by collecting multiple phone and email/text messaging contact information for participants and reviewing this information at each visit. It is possible that some participants may not have access to the internet or have technical challenges with Zoom. We will provide iPads and internet access for participants who lack equipment and/or internet access. In addition, our study team will teach participants how to use Zoom as needed and be available in real time to address technical issues. To note, synchronous online platforms have successfully been used to administer group-based psychobehavioral interventions, including in older populations [116–119]. Further, we have experience successfully conducting research groups via Zoom (Saban: PCORI). Despite these limitations, the proposed study will be designed and implemented with the highest degree of scientific rigor and has the potential for significant public health impact. Finally, by providing concrete recommendations based on the results of this intervention, we believe that this is the first study to provide the needed science to implement interventions focused on reducing the impact of multiple forms of stressors on the CMD health of AA women by exploring the biological changes in DNAm and inflammation that may contribute to this disparity.

Future directions

Lifestyle interventions can take 24 months to impact CMD risk [120] so likely would not be able to capture improvements in CMD risk indices in this proposed study. RiSE may benefit from larger epidemiological samples, other minority populations, and more frequent biological sample collection to detect changes in DNAm and inflammatory burden over longer periods of time. Future studies should evaluate the long-term impact of RiSE on CMD as well as on other inflammatory-related disease outcomes.

Author Contributions

Conceptualization: Jacquelyn Y. Taylor, Karen Saban.

Funding acquisition: Jacquelyn Y. Taylor, Karen Saban.

Investigation: Jacquelyn Y. Taylor, Karen Saban.

Methodology: Jacquelyn Y. Taylor, Karen Saban.

Project administration: Jacquelyn Y. Taylor, Karen Saban.

Resources: Jacquelyn Y. Taylor, Karen Saban.

Supervision: Jacquelyn Y. Taylor, Karen Saban.

Visualization: Jacquelyn Y. Taylor, Karen Saban.

Writing – original draft: Jacquelyn Y. Taylor, Alexandria Jones-Patten, Laura Prescott, Stephanie Potts-Thompson, Cara Joyce, Bamidele Tayo, Karen Saban.

Writing – review & editing: Jacquelyn Y. Taylor, Alexandria Jones-Patten, Laura Prescott, Stephanie Potts-Thompson, Cara Joyce, Bamidele Tayo, Karen Saban.

References

1. Chmielewski M, Lindholm B, Stenvinkel P, Ekström JT. The role of epigenetics in kidney diseases. *Prilozi*. 2011; 32(1):45–54. PMID: [21822177](#).
2. Bayarsaihan D. Epigenetic mechanisms in inflammation. *J Dent Res*. 2011 Jan; 90(1):9–17. <https://doi.org/10.1177/0022034510378683> PMID: [21178119](#); PMCID: PMC3144097.
3. Davis FM, Gallagher KA. Epigenetic Mechanisms in Monocytes/Macrophages Regulate Inflammation in Cardiometabolic and Vascular Disease. *Arterioscler Thromb Vasc Biol*. 2019 Apr; 39(4):623–634. <https://doi.org/10.1161/ATVBAHA.118.312135> PMID: [30760015](#); PMCID: PMC6438376.
4. Costantino S, Mohammed SA, Ambrosini S, Paneni F. Epigenetic processing in cardiometabolic disease. *Atherosclerosis*. 2019 Feb; 281:150–158. <https://doi.org/10.1016/j.atherosclerosis.2018.09.029> Epub 2018 Sep 26. PMID: [30290963](#).
5. Mendelson MM, Marioni RE, Joehanes R, Liu C, Hedman ÅK, Aslibekyan S, et al. Association of Body Mass Index with DNA Methylation and Gene Expression in Blood Cells and Relations to Cardiometabolic Disease: A Mendelian Randomization Approach. *PLoS Med*. 2017 Jan 17; 14(1): e1002215. <https://doi.org/10.1371/journal.pmed.1002215> PMID: [28095459](#); PMCID: PMC5240936.
6. Fiacco S, Giardini ES, Mernone L, Schick L, Ehler U. DNA Methylation in Healthy Older Adults with a History of Childhood Adversity-Findings From the Women 40+ Healthy Aging Study. *Front Psychiatry*. 2019 Oct 23; 10:777. <https://doi.org/10.3389/fpsy.2019.00777> PMID: [31708823](#); PMCID: PMC6819958.
7. Kundakovic M, Gudsnuk K, Herbstman JB, Tang D, Perera FP, Champagne FA. DNA methylation of BDNF as a biomarker of early-life adversity. *Proc Natl Acad Sci U S A*. 2015 Jun 2; 112(22):6807–13. <https://doi.org/10.1073/pnas.1408355111> Epub 2014 Nov 10. PMID: [25385582](#); PMCID: PMC4460453.
8. Turecki G, Meaney MJ. Effects of the Social Environment and Stress on Glucocorticoid Receptor Gene Methylation: A Systematic Review. *Biol Psychiatry*. 2016 Jan 15; 79(2):87–96. <https://doi.org/10.1016/j.biopsych.2014.11.022> Epub 2014 Dec 13. PMID: [25687413](#); PMCID: PMC4466091.
9. Mulder RH, Walton E, Neumann A, Houtepen LC, Felix JF, Bakermans-Kranenburg MJ, et al. Epigenomics of being bullied: changes in DNA methylation following bullying exposure. *Epigenetics*. 2020 Jun-Jul; 15(6–7):750–764. <https://doi.org/10.1080/15592294.2020.1719303> Epub 2020 Jan 28. PMID: [31992121](#); PMCID: PMC7574379.
10. Kang JI, Kim TY, Choi JH, So HS, Kim SJ. The allele-specific DNA methylation level of FKBP5 is associated with post-traumatic stress disorder. *Psychoneuroendocrinology*. 2019 May; 103:1–7. <https://doi.org/10.1016/j.psyneuen.2018.12.226> Epub 2018 Dec 19. PMID: [30605803](#).
11. Matosin N, Cruceanu C, Binder EB. Preclinical and Clinical Evidence of DNA Methylation Changes in Response to Trauma and Chronic Stress. *Chronic Stress (Thousand Oaks)*. 2017 Feb 1; 1:2470547017710764. <https://doi.org/10.1177/2470547017710764> Epub 2017 Jun 16. PMID: [29503977](#); PMCID: PMC5831952.
12. Barcelona de Mendoza V, Huang Y, Crusto CA, Sun YV, Taylor JY. Perceived Racial Discrimination and DNA Methylation Among African American Women in the InterGEN Study. *Biol Res Nurs*. 2018 Mar; 20(2):145–152. <https://doi.org/10.1177/1099800417748759> Epub 2017 Dec 19. PMID: [29258399](#); PMCID: PMC5741522.
13. Szyf M, Tang YY, Hill KG, Musci R. The dynamic epigenome and its implications for behavioral interventions: a role for epigenetics to inform disorder prevention and health promotion. *Transl Behav Med*. 2016; 6(1):55–62. <https://doi.org/10.1007/s13142-016-0387-7> PMID: [27012253](#)
14. Stoffel M, Gardini E, Ehrental JC, Abbruzzese E, Ditzen B. Evaluation of stress management and stress prevention using epigenetic markers. Review. *Verhaltenstherapie*. 2020. 30(1):18–27. <https://doi.org/10.1159/000505595>
15. McBride CM, Koehly LM. Imagining roles for epigenetics in health promotion research. *J Behav Med*. 2017 Apr; 40(2):229–238. <https://doi.org/10.1007/s10865-016-9764-4> Epub 2016 Jul 13. PMID: [27412775](#); PMCID: PMC5332486.

16. Hoye JR, Cheishvili D, Yarger HA, Roth TL, Szyf M, Dozier M. Preliminary indications that the Attachment and Biobehavioral Catch-up Intervention alters DNA methylation in maltreated children. *Dev Psychopathol*. 2020 Oct; 32(4):1486–1494. <https://doi.org/10.1017/S0954579419001421> PMID: 31854285.
17. Bishop JR, Lee AM, Mills LJ, Thuras PD, Eum S, Clancy D, et al. Methylation of FKBP5 and SLC6A4 in Relation to Treatment Response to Mindfulness Based Stress Reduction for Posttraumatic Stress Disorder. *Frontiers in Psychiatry*. 2018 Sep 18; 9:418. <https://doi.org/10.3389/fpsy.2018.00418> PMID: 30279666
18. Bishop JR, Lee AM, Mills LJ, Thuras PD, Eum S, Clancy D, et al. Corrigendum: Methylation of FKBP5 and SLC6A4 in Relation to Treatment Response to Mindfulness Based Stress Reduction for Posttraumatic Stress Disorder. *Front Psychiatry*. 2018 Sep 18; 9:418. <https://doi.org/10.3389/fpsy.2018.00418> Erratum in: *Front Psychiatry*. 2021 Mar 04; 12:642245. PMID: 30279666; PMCID: PMC6153325.
19. Stoffel M, Aguilar-Raab C, Rahn S, Steinhilber B, Witt SH, Alexander N, et al. Effects of Mindfulness-Based Stress Prevention on Serotonin Transporter Gene Methylation. *Psychother Psychosom*. 2019; 88(5):317–319. <https://doi.org/10.1159/000501646> Epub 2019 Aug 28. PMID: 31461722.
20. Harkess KN, Ryan J, Delfabbro PH, Cohen-Woods S. Preliminary indications of the effect of a brief yoga intervention on markers of inflammation and DNA methylation in chronically stressed women. *Transl Psychiatry*. 2016 Nov 29; 6(11): e965. <https://doi.org/10.1038/tp.2016.234> PMID: 27898068; PMCID: PMC5290356.
21. Ren H, Collins V, Clarke SJ, Han JS, Lam P, Clay F, et al. Epigenetic changes in response to tai chi practice: a pilot investigation of DNA methylation marks. *Evid Based Complement Alternat Med*. 2012; 2012:841810. <https://doi.org/10.1155/2012/841810> Epub 2012 Jun 5. PMID: 22719790; PMCID: PMC3375016.
22. Yehuda R, Daskalakis NP, Desarnaud F, Makotkine I, Lehrner AL, Koch E, et al. Epigenetic Biomarkers as Predictors and Correlates of Symptom Improvement Following Psychotherapy in Combat Veterans with PTSD. *Front Psychiatry*. 2013 Sep 27; 4:118. <https://doi.org/10.3389/fpsy.2013.00118> PMID: 24098286; PMCID: PMC3784793.
23. Chaix R, Fagny M, Cosin-Tomás M, Alvarez-López M, Lemee L, Regnault B, et al. Differential DNA methylation in experienced meditators after an intensive day of mindfulness-based practice: Implications for immune-related pathways. *Brain Behav Immun*. 2020 Feb; 84:36–44. <https://doi.org/10.1016/j.bbi.2019.11.003> Epub 2019 Nov 13. PMID: 31733290; PMCID: PMC7010561.
24. Roberts S, Keers R, Breen G, Coleman JRI, Jöhren P, Kepa A, et al. DNA methylation of FKBP5 and response to exposure-based psychological therapy. *Am J Med Genet B Neuropsychiatr Genet*. 2019 Mar; 180(2):150–158. <https://doi.org/10.1002/ajmg.b.32650> Epub 2018 Oct 18. PMID: 30334356; PMCID: PMC6600698.
25. Perroud N, Salzmann A, Prada P, Nicastro R, Hoeppli ME, Furrer S, et al. Response to psychotherapy in borderline personality disorder and methylation status of the BDNF gene. *Transl Psychiatry*. 2013 Jan 15; 3(1): e207. <https://doi.org/10.1038/tp.2012.140> PMID: 23422958; PMCID: PMC3566720.
26. Thomas M, Knoblich N, Wallisch A, Glowacz K, Becker-Sadzio J, Gundel F, et al. Increased BDNF methylation in saliva, but not blood, of patients with borderline personality disorder. *Clin Epigenetics*. 2018 Aug 22; 10(1):109. <https://doi.org/10.1186/s13148-018-0544-6> PMID: 30134995; PMCID: PMC6106893.
27. Kessler T, Vilne B, Schunkert H. The impact of genome-wide association studies on the pathophysiology and therapy of cardiovascular disease. *EMBO Mol Med*. 2016 Jul 1; 8(7):688–701. <https://doi.org/10.15252/emmm.201506174> PMID: 27189168; PMCID: PMC4931285.
28. Taylor JY, Schwander K, Kardina SL, Arnett D, Liang J, Hunt SC, et al. A Genome-wide study of blood pressure in African Americans accounting for gene-smoking interaction. *Sci Rep*. 2016 Jan 11; 6:18812. <https://doi.org/10.1038/srep18812> PMID: 26752167; PMCID: PMC4707536.
29. Wagner J, Abbott G. Depression and depression care in diabetes: relationship to perceived discrimination in African Americans. *Diabetes Care*. 2007 Feb; 30(2):364–6. <https://doi.org/10.2337/dc06-1756> PMID: 17259510.
30. Barber S, Hickson DA, Kawachi I, Subramanian SV, Earls F. Neighborhood Disadvantage and Cumulative Biological Risk Among a Socioeconomically Diverse Sample of African American Adults: An Examination in the Jackson Heart Study. *J Racial Ethn Health Disparities*. 2016 Sep; 3(3):444–56. <https://doi.org/10.1007/s40615-015-0157-0> Epub 2015 Sep 28. PMID: 27294737; PMCID: PMC4911317.
31. Belgrave FZ, Abrams JA. Reducing disparities and achieving equity in African American women's health. *Am Psychol*. 2016 Nov; 71(8):723–733. <https://doi.org/10.1037/amp0000081> PMID: 27977253.

32. Kershaw KN, Lewis TT, Diez Roux AV, Jenny NS, Liu K, Penedo FJ, et al. Self-reported experiences of discrimination and inflammation among men and women: The multi-ethnic study of atherosclerosis. *Health Psychol*. 2016 Apr; 35(4):343–50. <https://doi.org/10.1037/hea0000331> PMID: 27018725; PMCID: PMC4817357.
33. Mouton CP, Hayden M, Southerland JH. Cardiovascular Health Disparities in Underserved Populations. *Prim Care*. 2017 Mar; 44(1): e37–e71. <https://doi.org/10.1016/j.pop.2016.09.019> PMID: 28164826.
34. Wagner J, Lampert R, Tennen H, Feinn R. Exposure to Discrimination and Heart Rate Variability Reactivity to Acute Stress among Women with Diabetes. *Stress Health*. 2015 Aug; 31(3):255–62. <https://doi.org/10.1002/smi.2542> Epub 2013 Nov 6. PMID: 24194397.
35. Mwendwa DT, Sims RC, Madhere S, Thomas J, Keen LD 3rd, Callender CO, et al. The influence of coping with perceived racism and stress on lipid levels in African Americans. *J Natl Med Assoc*. 2011 Jul; 103(7):594–601. [https://doi.org/10.1016/s0027-9684\(15\)30385-0](https://doi.org/10.1016/s0027-9684(15)30385-0) PMID: 21999034; PMCID: PMC5003038.
36. Lewis JA, Williams MG, Peppers EJ, Gadson CA. Applying intersectionality to explore the relations between gendered racism and health among Black women. *J Couns Psychol*. 2017 Oct; 64(5):475–486. <https://doi.org/10.1037/cou0000231> PMID: 29048194.
37. Carlson M., Endlsey M., Motley D., Shawahin L. N., & Williams M. T. Addressing the impact of racism on veterans of color: A race-based stress and trauma intervention. *Psychology of Violence*. 2018. 8 (6), 748–762. <https://doi.org/10.1037/vio0000221>
38. Saban KL, Motley D, Shawahin L, Mathews HL, Tell D, De La Pena P, et al. Preliminary evidence for a race-based stress reduction intervention for Black women at risk for cardiovascular disease. *Complement Ther Med*. 2021 May; 58:102710. <https://doi.org/10.1016/j.ctim.2021.102710> Epub 2021 Mar 13. PMID: 33727090.
39. Motley D, Shawahin L. Race Based Stress and Resilience Group. Clinical Intervention Handbook. Veterans Affairs. 2017.
40. Beck J. Cognitive behavior therapy: Basics and beyond. Guilford Press. 2011.
41. Cokley Kevin & Hall-Clark Brittany & Hicks Dana. Ethnic Minority-Majority Status and Mental Health: The Mediating Role of Perceived Discrimination. *Journal of Mental Health Counseling*. 2011. 33. 243–263. <https://doi.org/10.17744/mehc.33.3.u1n011t020783086>
42. Pascoe EA, Smart Richman L. Perceived discrimination and health: a meta-analytic review. *Psychol Bull*. 2009 Jul; 135(4):531–54. <https://doi.org/10.1037/a0016059> PMID: 19586161; PMCID: PMC2747726.
43. Helms JE. Black and White Racial Identity: Theory, research, and practice. Greenwood Press. 1990.
44. Nieto L. Beyond inclusion, beyond empowerment: A Developmental strategy to liberate everyone. *Cuetzpalin*. 2010.
45. Williams MT, Chapman LK, Wong J, Turkheimer E. The role of ethnic identity in symptoms of anxiety and depression in African Americans. *Psychiatry Res*. 2012 Aug 30; 199(1):31–6. <https://doi.org/10.1016/j.psychres.2012.03.049> Epub 2012 Apr 17. PMID: 22513043; PMCID: PMC3445759.
46. Morrow S. L., & Hawxhurst D. M. Feminist therapy: Integrating political analysis in counseling and psychotherapy. *Women & Therapy*. 1998. 21(2), 37–50. https://doi.org/10.1300/J015v21n02_03
47. McEwen BS. Stress, adaptation, and disease. Allostasis and allostatic load. *Ann N Y Acad Sci*. 1998 May 1; 840: 33–44. <https://doi.org/10.1111/j.1749-6632.1998.tb09546.x> PMID: 9629234.
48. Conway-Phillips R, Dagadu H, Motley D, Shawahin L, Janusek LW, Klonowski S, et al. Qualitative evidence for Resilience, Stress, and Ethnicity (RiSE): A program to address race-based stress among Black women at risk for cardiovascular disease. *Complement Ther Med*. 2020 Jan; 48:102277. <https://doi.org/10.1016/j.ctim.2019.102277> Epub 2019 Dec 12. PMID: 31987226.
49. Brown KM, Hui Q, Huang Y, Taylor JY, Prescott L, de Mendoza VB, et al. Association Between Stress and Coping with DNA Methylation of Blood Pressure-Related Genes Among African American Women. *Chronic Stress (Thousand Oaks)*. 2019 Jan-Dec; 3:2470547019879088. <https://doi.org/10.1177/2470547019879088> Epub 2019 Sep 26. PMID: 32395678; PMCID: PMC7213592.
50. Bukowska-Damska A, Reszka E, Kaluzny P, Wiecek E, Przybek M, Zienoldiny S, et al. Sleep quality and methylation status of core circadian rhythm genes among nurses and midwives. *Chronobiol Int*. 2017; 34(9):1211–1223. <https://doi.org/10.1080/07420528.2017.1358176> Epub 2017 Nov 6. PMID: 29106308.
51. Carskadon MA, Chappell KR, Barker DH, Hart AC, Dwyer K, Gredvig-Ardito C, et al. A pilot prospective study of sleep patterns and DNA methylation-characterized epigenetic aging in young adults. *BMC Res Notes*. 2019 Sep 16; 12(1):583. <https://doi.org/10.1186/s13104-019-4633-1> PMID: 31526398; PMCID: PMC6747743.

52. Glad CA, Andersson-Assarsson JC, Berglund P, Bergthorsdottir R, Ragnarsson O, Johannsson G. Reduced DNA methylation and psychopathology following endogenous hypercortisolism—a genome-wide study. *Sci Rep*. 2017 Mar 16; 7:44445. <https://doi.org/10.1038/srep44445> PMID: 28300138; PMCID: PMC5353706.
53. Janusek LW, Tell D, Gaylord-Harden N, Mathews HL. Relationship of childhood adversity and neighborhood violence to a proinflammatory phenotype in emerging adult African American men: An epigenetic link. *Brain Behav Immun*. 2017 Feb; 60:126–135. <https://doi.org/10.1016/j.bbi.2016.10.006> Epub 2016 Oct 17. PMID: 27765646.
54. Walsh E, Eisenlohr-Moul T, Baer R. Brief mindfulness training reduces salivary IL-6 and TNF- α in young women with depressive symptomatology. *J Consult Clin Psychol*. 2016 Oct; 84(10):887–97. <https://doi.org/10.1037/ccp0000122> Epub 2016 Jun 9. PMID: 27281371; PMCID: PMC5037002.
55. Lengacher CA, Reich RR, Paterson CL, et al. A Large Randomized Trial: Effects of Mindfulness-Based Stress Reduction (MBSR) for Breast Cancer (BC) Survivors on Salivary Cortisol and IL-6. *Biol Res Nurs*. Jan 2019; 21(1):39–49. <https://doi.org/10.1177/1099800418789777> PMID: 30079756
56. Womersley J. S., Nothling J., Toikumo S., Malan-Müller S., van den Heuvel L. L., McGregor N. W., et al., (2022). Childhood trauma, the stress response and metabolic syndrome: A focus on DNA methylation. *The European journal of neuroscience*, 55(9–10), 2253–2296. <https://doi.org/10.1111/ejn.15370> PMID: 34169602
57. Martelle SE, Cotella EM, Nawreen N, Chen C, Packard BA, Fitzgerald M, et al. Prefrontal cortex PACAP signaling: organization and role in stress regulation. *Stress*. 2021 Mar; 24(2):196–205. <https://doi.org/10.1080/10253890.2021.1887849> Epub 2021 Mar 17. PMID: 33726625; PMCID: PMC8025233.
58. Swartz JR, Hariri AR, Williamson DE. An epigenetic mechanism links socioeconomic status to changes in depression-related brain function in high-risk adolescents. *Molecular Psychiatry*. 2017/02/01 2017; 22(2):209–214. <https://doi.org/10.1038/mp.2016.82> PMID: 27217150
59. Kobiella A, Reimold M, Ulshöfer DE, Ikonomidou VN, Vollmert C, Vollstädt-Klein S, et al. How the serotonin transporter 5-HTTLPR polymorphism influences amygdala function: the roles of in vivo serotonin transporter expression and amygdala structure. *Transl Psychiatry*. 2011 Aug 30; 1(8): e37. <https://doi.org/10.1038/tp.2011.29> PMID: 22832611; PMCID: PMC3309509.
60. Miao Z, Wang Y, Sun Z. The Relationships Between Stress, Mental Disorders, and Epigenetic Regulation of BDNF. *International journal of molecular sciences*. 2020; 21(4):1375. <https://doi.org/10.3390/ijms21041375> PMID: 32085670
61. Pillai A, Bruno D, Sarreal AS, Hernando RT, Saint-Louis LA, Nierenberg J, et al. Plasma BDNF levels vary in relation to body weight in females. *PLoS One*. 2012; 7(7): e39358. <https://doi.org/10.1371/journal.pone.0039358> Epub 2012 Jul 2. PMID: 22768299; PMCID: PMC3388065.
62. Smith JA, Zhao W, Wang X, Ratliff SM, Mukherjee B, Kardias SLR, et al. Neighborhood characteristics influence DNA methylation of genes involved in stress response and inflammation: The Multi-Ethnic Study of Atherosclerosis. *Epigenetics*. 2017 Aug; 12(8):662–673. <https://doi.org/10.1080/15592294.2017.1341026> Epub 2017 Jul 5. PMID: 28678593; PMCID: PMC5687339.
63. Provenzi L, Brambilla M, Scotto di Minico G, Montirosso R, Borgatti R. Maternal caregiving and DNA methylation in human infants and children: Systematic review. *Genes Brain Behav*. Mar 2020; 19(3): e12616. <https://doi.org/10.1111/gbb.12616> PMID: 31622002
64. Kogan SM, Bae D, Cho J, Smith AK, Nishitani S. Pathways linking adverse environments to emerging adults' substance abuse and depressive symptoms: A prospective analysis of rural African American men. *Dev Psychopathol*. Jul 22, 2020:1–11. <https://doi.org/10.1017/s0954579420000632> PMID: 32693849
65. Ursini G, Bollati V, Fazio L, Porcelli A, Iacovelli L, Catalani A, et al. Stress-related methylation of the catechol-O-methyltransferase Val 158 allele predicts human prefrontal cognition and activity. *J Neurosci*. 2011 May 4; 31(18):6692–8. <https://doi.org/10.1523/JNEUROSCI.6631-10.2011> PMID: 21543598; PMCID: PMC6632869.
66. Luo Y, Zheng SG. Hall of Fame among Pro-inflammatory Cytokines: Interleukin-6 Gene and Its Transcriptional Regulation Mechanisms. Review. *Frontiers in Immunology*. 2016-December-19 2016; 7(604) <https://doi.org/10.3389/fimmu.2016.00604> PMID: 28066415
67. Wendt FR, Pathak GA, Levey DF, Nuñez YZ, Overstreet C, Tyrrell C, et al. Sex-stratified gene-by-environment genome-wide interaction study of trauma, posttraumatic-stress, and suicidality. *Neurobiol Stress*. 2021 Feb 18; 14:100309. <https://doi.org/10.1016/j.ynstr.2021.100309> PMID: 33665242; PMCID: PMC7905234.
68. Bakusic J, Vrieze E, Ghosh M, Bekaert B, Claes S, Godderis L. Increased methylation of NR3C1 and SLC6A4 is associated with blunted cortisol reactivity to stress in major depression. *Neurobiol Stress*. Nov 2020; 13:100272. <https://doi.org/10.1016/j.ynstr.2020.100272> PMID: 33344725

69. Santos HP Jr, Adynski H, Harris R, Bhattacharya A, Incollingo Rodriguez AC, et al. Biopsychosocial correlates of psychological distress in Latina mothers. *J Affect Disord*. 2021 Mar 1; 282:617–626. <https://doi.org/10.1016/j.jad.2020.12.193> Epub 2020 Dec 31. PMID: 33445084; PMCID: PMC7889736.
70. Hao G, Youssef NA, Davis CL, Su S. The role of DNA methylation in the association between childhood adversity and cardiometabolic disease. *Int J Cardiol*. Mar 15 2018; 255:168–174. <https://doi.org/10.1016/j.ijcard.2017.12.063> PMID: 29288057
71. Houtepen LC, Vinkers CH, Carrillo-Roa T, Hiemstra M, van Lier PA, Meeus W, et al. Genome-wide DNA methylation levels and altered cortisol stress reactivity following childhood trauma in humans. *Nat Commun*. 2016 Mar 21; 7:10967. <https://doi.org/10.1038/ncomms10967> PMID: 26997371; PMCID: PMC4802173.
72. Jorgensen BG, Berent RM, Ha SE, Horiguchi K, Sasse KC, Becker LS, et al. DNA methylation, through DNMT1, has an essential role in the development of gastrointestinal smooth muscle cells and disease. *Cell Death Dis*. 2018 May 1; 9(5):474. <https://doi.org/10.1038/s41419-018-0495-z> PMID: 29700293; PMCID: PMC5920081.
73. Vidrascu EM, Bashore AC, Howard TD, Moore JB. Effects of early- and mid-life stress on DNA methylation of genes associated with subclinical cardiovascular disease and cognitive impairment: a systematic review. *BMC medical genetics*. 2019/03/12 2019; 20(1):39. <https://doi.org/10.1186/s12881-019-0764-4> PMID: 30866842
74. Phumsatitpong C, Wagenmaker ER, Moenter SM. Neuroendocrine interactions of the stress and reproductive axes. *Front Neuroendocrinol*. Jun 22, 2021:100928. <https://doi.org/10.1016/j.yfrne.2021.100928> PMID: 34171353
75. Friso S, Pizzolo F, Choi SW, Guarini P, Castagna A, Ravagnani V, et al. Epigenetic control of 11 beta-hydroxysteroid dehydrogenase 2 gene promoter is related to human hypertension. *Atherosclerosis*. 2008 Aug; 199(2):323–7. <https://doi.org/10.1016/j.atherosclerosis.2007.11.029> Epub 2008 Feb 7. PMID: 18178212.
76. Argentieri MA, Nagarajan S, Seddighzadeh B, Baccarelli AA, Shields AE. Epigenetic Pathways in Human Disease: The Impact of DNA Methylation on Stress-Related Pathogenesis and Current Challenges in Biomarker Development. *EBioMedicine*. Apr 2017; 18:327–350. <https://doi.org/10.1016/j.ebiom.2017.03.044> PMID: 28434943
77. Sun X, Ming Q, Zhong X, Dong D, Li C, Xiong G, et al. The MAOA Gene Influences the Neural Response to Psychosocial Stress in the Human Brain. *Front Behav Neurosci*. 2020 May 15; 14:65. <https://doi.org/10.3389/fnbeh.2020.00065> PMID: 32499684; PMCID: PMC7243356.
78. Zill P, Baghai TC, Schüle C, Born C, Früstück C, Büttner A, et al. DNA methylation analysis of the angiotensin converting enzyme (ACE) gene in major depression. *PLoS One*. 2012; 7(7): e40479. <https://doi.org/10.1371/journal.pone.0040479> Epub 2012 Jul 13. PMID: 22808171; PMCID: PMC3396656.
79. Lindholm ME, Marabita F, Gomez-Cabrero D, Rundqvist H, Ekström TJ, Tegnér J, et al. An integrative analysis reveals coordinated reprogramming of the epigenome and the transcriptome in human skeletal muscle after training. *Epigenetics*. 2014 Dec; 9(12):1557–69. <https://doi.org/10.4161/15592294.2014.982445> PMID: 25484259; PMCID: PMC4622000.
80. Zhang Y, Hashimoto S, Fujii C, et al. NFκB2 Gene as a Novel Candidate that Epigenetically Responds to Interval Walking Training. *International journal of sports medicine*. Aug 2015; 36(9):769–75. <https://doi.org/10.1055/s-0035-1547221> PMID: 25901949
81. Tsaprouni LG, Yang TP, Bell J, Dick KJ, Kanoni S, Nisbet J, et al. Cigarette smoking reduces DNA methylation levels at multiple genomic loci but the effect is partially reversible upon cessation. *Epigenetics*. 2014 Oct; 9(10):1382–96. <https://doi.org/10.4161/15592294.2014.969637> PMID: 25424692; PMCID: PMC4623553.
82. Nelson KN, Hui Q, Rimland D, et al. Identification of HIV infection-related DNA methylation sites and advanced epigenetic aging in HIV-positive, treatment-naive U.S. veterans. *Aids*. Feb 20, 2017; 31(4):571–575. <https://doi.org/10.1097/QAD.0000000000001360> PMID: 27922854
83. Dantzer R, Cohen S, Russo SJ, Dinan TG. Resilience and immunity. *Brain Behav Immun*. Nov 2018; 74:28–42. <https://doi.org/10.1016/j.bbi.2018.08.010> PMID: 30102966
84. Elenkov IJ, Chrousos GP. Stress hormones, proinflammatory and antiinflammatory cytokines, and autoimmunity. *Ann N Y Acad Sci*. Jun 2002; 966:290–303. <https://doi.org/10.1111/j.1749-6632.2002.tb04229.x> PMID: 12114286
85. Paudel D, Morikawa T, Yoshida K, Uehara O, Giri S, Neopane P, et al. Chronic stress-induced elevation of IL-1β in the saliva and submandibular glands of mice. *Med Mol Morphol*. 2020 Dec; 53(4):238–243. <https://doi.org/10.1007/s00795-020-00250-w> Epub 2020 Apr 6. PMID: 32253605.

86. Takemori Y, Sasayama D, Toida Y, Kotagiri M, Sugiyama N, Yamaguchi M, et al. Possible utilization of salivary IFN- γ /IL-4 ratio as a marker of chronic stress in healthy individuals. *Neuropsychopharmacol Rep*. 2021 Mar; 41(1):65–72. <https://doi.org/10.1002/npr2.12157> Epub 2021 Jan 19. PMID: 33465301; PMCID: PMC8182956.
87. Szabo YZ, Slavish DC, Graham-Engeland JE. The effect of acute stress on salivary markers of inflammation: A systematic review and meta-analysis. *Brain Behav Immun*. Aug 2020; 88:887–900. <https://doi.org/10.1016/j.bbi.2020.04.078> PMID: 32371089
88. Slavish DC, Szabo YZ. What moderates salivary markers of inflammation reactivity to stress? A descriptive report and meta-regression. *Stress*. Mar 24, 2021:1–13. <https://doi.org/10.1080/10253890.2021.1887848> PMID: 33759687
89. Siddiqui A, Madhu SV, Sharma SB, Desai NG. Endocrine stress responses and risk of type 2 diabetes mellitus. *Stress*. 2015; 18(5):498–506. <https://doi.org/10.3109/10253890.2015.1067677> PMID: 26303379
90. Saban K, Jones-Patten A, Janusek L, Tell D, de la Pena P, Prescott L, et al. 2023. Race-Based Stress Reduction Intervention (RiSE) study in Chicago and NYC: Design and Methods for Recruitment and Intervention. *PLoS One*. Forthcoming.
91. Guo F, Moellering DR, Garvey WT. The progression of cardiometabolic disease: validation of a new cardiometabolic disease staging system applicable to obesity. *Obesity* (Silver Spring, Md). Jan 2014; 22(1):110–8. <https://doi.org/10.1002/oby.20585> PMID: 23894121
92. Riis JL, Ahmadi H, Hamilton KR, Hand T, Granger DA. Best practice recommendations for the measurement and interpretation of salivary proinflammatory cytokines in biobehavioral research. *Brain Behav Immun*. 2021 Jan; 91:105–116. <https://doi.org/10.1016/j.bbi.2020.09.009> Epub 2020 Sep 12. PMID: 32931871; PMCID: PMC8164445.
93. Morrison-Beedy D, Carey MP, Tu X. Accuracy of audio computer-assisted self-interviewing (ACASI) and self-administered questionnaires for the assessment of sexual behavior. *AIDS Behav*. 2006; 10(5):541–552. <https://doi.org/10.1007/s10461-006-9081-y> PMID: 16721506
94. Cohen S, Kamarck T, Mermelstein R. A global measure of perceived stress. *J Health Soc Behav*. 1983 Dec; 24(4):385–96. PMID: 6668417.
95. Kroenke K, Spitzer RL, Williams JB. The PHQ-9: validity of a brief depression severity measure. *J Gen Intern Med*. 2001 Sep; 16(9):606–13. <https://doi.org/10.1046/j.1525-1497.2001.016009606.x> PMID: 11556941; PMCID: PMC1495268.
96. HealthMeasures. PROMIS 2016. <http://www.nihpromis.org/>
97. Morin C. *Insomnia: Psychological Assessment and Management*. Guilford Press. 1993
98. Thompson TM, Sharfi D, Lee M, Yrigollen CM, Naumova OY, Grigorenko EL. Comparison of whole-genome DNA methylation patterns in whole blood, saliva, and lymphoblastoid cell lines. *Behav Genet*. 2013 Mar; 43(2):168–76. <https://doi.org/10.1007/s10519-012-9579-1> Epub 2012 Dec 27. PMID: 23269419; PMCID: PMC3577999.
99. Langie SAS, Moisse M, Declerck K, Koppen G, Godderis L, Vanden Berghe W, et al. Salivary DNA Methylation Profiling: Aspects to Consider for Biomarker Identification. *Basic Clin Pharmacol Toxicol*. 2017 Sep; 121 Suppl 3 (Suppl 3):93–101. <https://doi.org/10.1111/bcpt.12721> Epub 2017 Feb 10. PMID: 27901320; PMCID: PMC5644718.
100. Langie SA, Szarc Vel Szic K, Declerck K, Traen S, Koppen G, et al. Whole-Genome Saliva and Blood DNA Methylation Profiling in Individuals with a Respiratory Allergy. *PLoS One*. 2016 Mar 21; 11(3): e0151109. <https://doi.org/10.1371/journal.pone.0151109> Erratum in: *PLoS One*. 2017 Aug 7; 12(8): e0183088. PMID: 26999364; PMCID: PMC4801358.
101. Smith AK, Kilaru V, Klengel T, Mercer KB, Bradley B, Conneely KN, et al. DNA extracted from saliva for methylation studies of psychiatric traits: evidence tissue specificity and relatedness to brain. *Am J Med Genet B Neuropsychiatr Genet*. 2015 Jan; 168B(1):36–44. <https://doi.org/10.1002/ajmg.b.32278> Epub 2014 Oct 29. PMID: 25355443; PMCID: PMC4610814.
102. Braun PR, Han S, Hing B, Nagahama Y, Gaul LN, Heinzman JT, et al. Genome-wide DNA methylation comparison between live human brain and peripheral tissues within individuals. *Transl Psychiatry*. 2019 Jan 31; 9(1):47. <https://doi.org/10.1038/s41398-019-0376-y> PMID: 30705257; PMCID: PMC6355837.
103. Rounge TB, Page CM, Lepistö M, Eilonen P, Andreassen BK, Weiderpass E. Genome-wide DNA methylation in saliva and body size of adolescent girls. *Epigenomics*. Nov 2016; 8(11):1495–1505. <https://doi.org/10.2217/epi-2016-0045> PMID: 27762626
104. Chiarella J, Schumann L, Pomares FB, Frodt T, Tozzi L, Nemoda Z, et al. DNA methylation differences in stress-related genes, functional connectivity and gray matter volume in depressed and healthy adolescents. *J Affect Disord*. 2020 Jun 15; 271:160–168. <https://doi.org/10.1016/j.jad.2020.03.062> Epub 2020 Apr 6. PMID: 32479312.

105. Ismaylova E, Lévesque ML, Pomares FB, Szyf M, Nemoda Z, Fahim C, et al. Serotonin transporter promoter methylation in peripheral cells and neural responses to negative stimuli: A study of adolescent monozygotic twins. *Transl Psychiatry*. 2018 Aug 8; 8(1):147. <https://doi.org/10.1038/s41398-018-0195-6> PMID: 30089832; PMCID: PMC6082838.
106. Taylor JY, Wright ML, Crusto CA, Sun YV. The Intergenerational Impact of Genetic and Psychological Factors on Blood Pressure (InterGEN) Study: Design and Methods for Complex DNA Analysis. *Biol Res Nurs*. Oct 2016; 18(5):521–30. <https://doi.org/10.1177/1099800416645399> PMID: 27118148
107. Noguera-Castells A, García-Prieto CA, Álvarez-Errico D, Esteller M. Validation of the new EPIC DNA methylation microarray (900K EPIC v2) for high-throughput profiling of the human DNA methylome. *Epigenetics*. 2023 Dec; 18(1):2185742. <https://doi.org/10.1080/15592294.2023.2185742> PMID: 36871255; PMCID: PMC9988339.
108. Moran S, Arribas C, Esteller M. Validation of a DNA methylation microarray for 850,000 CpG sites of the human genome enriched in enhancer sequences. *Epigenomics*. 2016 Mar; 8(3):389–99. <https://doi.org/10.2217/epi.15.114> Epub 2015 Dec 17. PMID: 26673039; PMCID: PMC4864062.
109. Bibikova M, Barnes B, Tsan C, Ho V, Klotzle B, Le JM, et al., High density DNA methylation array with single CpG site resolution. *Genomics*. 2011 Oct; 98(4):288–95. <https://doi.org/10.1016/j.ygeno.2011.07.007> Epub 2011 Aug 2. PMID: 21839163.
110. Herman JG, Graff JR, Myöhänen S, Nelkin BD, Baylin SB. Methylation-specific PCR: a novel PCR assay for methylation status of CpG islands. *Proc Natl Acad Sci U S A*. 1996 Sep 3; 93(18):9821–6. <https://doi.org/10.1073/pnas.93.18.9821> PMID: 8790415; PMCID: PMC38513.
111. Zinn RL, Pruitt K, Eguchi S, Baylin SB, Herman JG. hTERT is expressed in cancer cell lines despite promoter DNA methylation by preservation of unmethylated DNA and active chromatin around the transcription start site. *Cancer Res*. Jan 1, 2007; 67(1):194–201. <https://doi.org/10.1158/0008-5472.CAN-06-3396> PMID: 17210699
112. Gonzalez A, Catherine N, Boyle M, et al. Healthy Foundations Study: a randomised controlled trial to evaluate biological embedding of early-life experiences. *BMJ Open*. Jan 26, 2018; 8(1): e018915. <https://doi.org/10.1136/bmjopen-2017-018915> PMID: 29374668
113. Graw S, Henn R, Thompson JA, Koestler DC. pwrEWAS: a user-friendly tool for comprehensive power estimation for epigenome wide association studies (EWAS). *BMC bioinformatics*. Apr 29, 2019; 20(1):218. <https://doi.org/10.1186/s12859-019-2804-7> PMID: 31035919
114. Vickers A. J., & Altman D. G. (2001). Statistics notes: Analysing controlled trials with baseline and follow up measurements. *BMJ (Clinical research ed.)*, 323(7321), 1123–1124. <https://doi.org/10.1136/bmj.323.7321.1123> PMID: 11701584
115. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res*. 2015 Apr 20; 43(7):e47. <https://doi.org/10.1093/nar/gkv007> Epub 2015 Jan 20. PMID: 25605792; PMCID: PMC4402510.
116. Jayewardene WP, Lohrmann DK, Erbe RG, Torabi MR. Effects of preventive online mindfulness interventions on stress and mindfulness: A meta-analysis of randomized controlled trials. *Preventive medicine reports*. Mar 2017; 5:150–159. <https://doi.org/10.1016/j.pmedr.2016.11.013> PMID: 28050336
117. Spijkerman MPJ, Pots WTM, Bohlmeijer ET. Effectiveness of online mindfulness-based interventions in improving mental health: A review and meta-analysis of randomised controlled trials. *Clinical Psychology Review*. 2016/04/01/ 2016; 45:102–114. <https://doi.org/10.1016/j.cpr.2016.03.009> PMID: 27111302
118. Milbury K., Kroll J., Chen A., Antonoff M. B., Snyder S., Higgins H., et al., (2021). Pilot Randomized Controlled Trial in Women with Non-Small Cell Lung Cancer to Assess the Feasibility of Delivering Group-Based Psychosocial Care via Videoconference. *Integrative cancer therapies*, 20, 15347354211052520. <https://doi.org/10.1177/15347354211052520> PMID: 34663123
119. Shapira S., Yeshua-Katz D., Cohn-Schwartz E., Aharonson-Daniel L., Sarid O., & Clarfield A. M. (2021). A pilot randomized controlled trial of a group intervention via Zoom to relieve loneliness and depressive symptoms among older persons during the COVID-19 outbreak. *Internet interventions*, 24, 100368. <https://doi.org/10.1016/j.invent.2021.100368> PMID: 33527072
120. Vetter M. L., Wadden T. A., Chittams J., Diewald L. K., Panigrahi E., Volger S., et al., (2013). Effect of lifestyle intervention on cardiometabolic risk factors: results of the POWER-UP trial. *International journal of obesity (2005)*, 37 Suppl 1(0 1), S19–S24. <https://doi.org/10.1038/ijo.2013.92> PMID: 23921777